Two new species of *Leptogium* (Collemataceae) with transversely septate ascospores from East Africa

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Abstract: Two epiphytic *Leptogium* species from East Africa, both with transversely septate ascospores, are here described as new. They produce plicate thalli and have a paraplectenchymatous proper exciple and a one cell layer thick cortex on the thalline exciple. *Leptogium bellum* has so far only been collected from Mt. Kilimanjaro, Tanzania, where it occurs from mid-montane forests up to the subalpine zone. *Leptogium tiinae* occurs in relatively open habitats at lower elevations, with the type specimen collected from the Taita Hills, Kenya. Additionally, details from a *Leptogium chloromeloides* isotype and a key for *Leptogium* species with transversely septate ascospores are provided.

Keywords: Eastern Arc, Mt. Kilimanjaro, Taita Hills, paleotropics, biodiversity hotspot, afromontane, *Leptogium* section *Leptogiopsis*, Peltigerales

INTRODUCTION

*Leptogium* (Ach.) Gray is the most species-rich genus within Collemataceae (Peltigerales), with its highest diversity in tropical regions (Otálora et al., 2014). While most *Leptogium* species have muriform spores, some species produce ascospores with only transverse septa and have traditionally been placed into *Leptogium* section *Leptogiopsis* Vain. Kitaura et al. (2013) list eleven such species: *Leptogium adpressum* Nyl., *Leptogium ankolense* C.W. Dodge, *Leptogium brebissonii* Mont., *Leptogium chloromeloides* Nyl., *Leptogium fusisporum* (Tuck.) C.W. Dodge, *Leptogium granadillae* (C.W. Dodge) Degel., *Leptogium longisporum* Kitaura & Marcelli, *Leptogium megapotamicum* Malme, *Leptogium pacificum* Vain., *Leptogium reticulatum* Nyl., and *Leptogium thoroldii* C.W. Dodge. Even though with similar ascospores, these species differ significantly in apothecial anatomy (Kitaura, 2012; Kitaura et al., 2013) and hence probably do not form a monophyletic group.

In their handbook of East African macrolichens, Swinscow and Krog (1988) confirm only one species of *Leptogium* with transversely septate ascospores, *L. adpressum*. Also *L. brebissonii* is mentioned, but only on the basis of sterile specimens. Some further species of this group have been reported from Africa in older literature, including *L. ankolense* from Uganda (Dodge, 1971), *L. thoroldii* from western Africa (Dodge, 1964), and *L. chloromeloides* from South Africa (Nylander, 1869).

We recently analyzed nearly 600 *Leptogium* specimens collected from Kenya and Tanzania (Kaasalainen et al., 2021). The results revealed that montane forests of the East African mountains, belonging to the Eastern Afromontane biodiversity hotspot, support a very high diversity of *Leptogium* species, most of them presently undescribed. Here we describe two such taxa, both with transversely septate ascospores.

MATERIAL AND METHODS

The material of Kaasalainen et al. (2021) were revisited and all fertile specimens with transversely septate ascospores were picked for closer inspection and compared with available older herbarium specimens. For details on the study region, sampling, and the phylogenetic analysis of the nuITS and mtSSU genetic regions of the specimens see Kaasalainen et al. (2021). The morphological and anatomical characters were assessed using a Leica S8APO stereo and an
Olympus BX51 compound light microscope, the latter equipped with a Deltapix Invenio 12EIII camera.

RESULTS AND DISCUSSIONS

In our material from East Africa, specimens with transversely septate ascospores occur only in two closely related taxa, OTUH1 and OTUH2 of Clade H (Fig. 4 in Kaasalainen et al., 2021). Closer inspection of the specimens revealed differences in morphology and anatomy, confirming that the two taxa represent distinct species, further characterized by contrasting altitudinal distributions. By comparing these specimens with herbarium material and published descriptions of all known Leptogium species with transversely septate ascospores, it became evident that both species are new to science.

Species of Leptogium with transversely septate ascospores are best distinguished based on anatomical differences in the apothecia, such as the presence or absence of a paraplectenchymatous proper exciple and cortical thickness of the thalline exciple (either consisting of only one or several cell layers), structure of outgrowths on the apothecial margin, and size of the ascospores. It should be emphasized that when measuring the length of ascospores in this group, only spores from mature apothecia (generally with flattened discs) should be analysed as the ascospores of young apothecia (generally with concave discs) tend to be shorter, wider and with fewer septa.

Both Leptogium bellum and L. tiinae have a thick paraplectenchymatous proper exciple but lack a paraplectenchymatous cortical exciple and apothecial base. Previously described Leptogium species with similar apothecial features include L. adpressum (Kitaura, 2012), L. longisporum (Kitaura et al., 2013), and L. thoroldii (Dodge, 1964). The diagnostic characteristics of these species are summarized in Table 1.

An additional species with a prominent paraplectenchymatous proper exciple is Leptogium granadillae, originally described from Costa Rica. However, L. granadillae also has a paraplectenchymatous cortex of thalline exciple (Dodge, 1933; Degelius, 1962), unlike the newly described species.

Table 1. Summary of the presently known Leptogium species with acicular, transversely septate ascospores, a paraplectenchymatous proper exciple, and thin (one-celled) cortex on the thalline exciple.

<table>
<thead>
<tr>
<th>Type</th>
<th>Known distribution</th>
<th>Apothecial margin</th>
<th>Ascospores (µm)</th>
<th>Comments</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Leptogium adpressum</td>
<td>H-NYL 41.254ttt</td>
<td>With braid-like resembling wrinkles.</td>
<td>50−60 × 7−8</td>
<td>Previous reports of this species from East Africa likely represent L. bellum and/or L. tiinae.</td>
<td>Nylander, 1858; Sierk, 1964; Swinscow and Krog, 1988; Kitaura, 2012.</td>
</tr>
<tr>
<td>Leptogium bellum sp. nov.</td>
<td>H 92463spv</td>
<td>With delicate sack-like nodules.</td>
<td>40−60 × 6−12</td>
<td></td>
<td>Kitaura et al., 2013.</td>
</tr>
<tr>
<td>Leptogium longisporum</td>
<td>SP</td>
<td>Densely ridged, cerebroid contorted.</td>
<td>55−90 × 5</td>
<td></td>
<td>Dodge, 1964.</td>
</tr>
<tr>
<td>Leptogium thoroldii</td>
<td>FH 00507796</td>
<td>Minutely and deeply scrobiculate.</td>
<td>50−55 × 3−4</td>
<td></td>
<td>Kitaura et al., 2021.</td>
</tr>
<tr>
<td>Leptogium tiinae sp. nov.</td>
<td>EA JR10017a (duplicate H 92463tttt)</td>
<td>With robust cerebroid structures.</td>
<td>60−90 × 5.9</td>
<td></td>
<td>* Type location</td>
</tr>
</tbody>
</table>

* Type location
Several of the *Leptogium* species with transversely septate ascospores are characterized by a presence of several cell layers of paraplectenchymatous tissue in the cortex of thalline exciple and a lack of paraplectenchymatous proper exciple. According to Kitaura (2012), the apothecia of *L. brebissonii* from Brazil have paraplectenchymatous cortex of thalline exciple and a collopletrenchymatous proper exciple. *Leptogium brebissonii* was originally described from the Canary Islands (Webb and Berthelot, 1840), and only sterile specimens have been reported from East Africa (Swinscow and Krog, 1988; Kaasalainen et al., 2021). In the phylogenetic analysis of Kaasalainen et al. (2021), such specimens grouped into Clade E and are thus not closely related to *L. bellum* and *L. tiinae*. Also *Leptogium ankolense* from Uganda (Dodge, 1971), *L. megapotamicum* from Brazil (Kitaura et al., 2013), and *L. pacificum* from the Philippines (Kitaura, 2012) are characterized by a several cell layers thick cortex of the thalline exciple and lack of paraplectenchymatous proper exciple. The last-mentioned taxon has also been reported to have a pseudocortex on its lower surface (Kitaura, 2012), which is uncharacteristic for *Leptogium* s. str. (Otálora et al., 2014).

*Leptogium chloromeloides*, on the other hand, seem to lack both apothecial para-plectenchymatous tissues. Even though the original description lacks the details of apothecial anatomy (Nylander, 1869) and the type specimen in DBN (Medeiros, 2019) was not available for loan, an isotype in G (G00047541, Afrique du Sud; Wood, J.M. 326) was most helpfully sent for inspection. The lobes of the specimen are up to 2 cm long and 5 mm wide, relatively thick, adnate, and rounded; margins are ascending, often clearly wrinkled, thicker and darker than the central parts of the thallus. The upper surface is light grey and slightly wrinkled on the central parts, with occasional ridges, darker especially towards the margins. The lower surface is longitudinally plicate, yellowish brown or grey. Isidia and phyllidia were not seen. Some pycnidia are present on the thickened margins of the thallus. Apothecia are mainly marginal, slightly congested, up to 1.9 mm in diam., with reddish brown, flat to convex discs. Apothecial margins are slightly wrinkled or uneven and without ornamentation. The cortex of thalline exciple is one cell layer thick and thick, paraplectenchymatous proper exciple is not present. Hymenium is 80–85 µm and subhymenium 50–55 µm thick (N=1). The few seen ascospores fit well the original description by Nylander (1869): 34–40 × 4.5 µm with 5–7 septa.

Additionally, two species lack the information of apothecial anatomy or have contradicting previous descriptions: *Leptogium reticulatum* from the Neotropics has a reticulately ridged thallus. We could not find mature apothecia in any of the available specimens (isotype H-NYL 41389, French Guiana; H-NYL 41390, Guadalupe; H-NYL 41391, Cuba). The original description also lacks information on ascospore and apothecial characters (Montagne, 1841). However, the thallus morphology of this species is quite distinct, and a sequence of *L. reticulatum* from Colombia did not group with our new species in the phylogenetic analysis (Kaasalainen et al., 2021). *Leptogium fusisporum* from the USA was originally described as having granular apothecial margins and ascospores up to 48 µm long and with 8–10 cells, but details of the apothecial anatomy were not given (Tuckerman, 1882; Dodge, 1933). According to Sierk (1964), the apothecia have entire margins, the proper exciple is paraplectenchymatous, the thalline exciple has a one layered cortex, and the ascospores are 25–40 × 6–7 µm with 3–5 septa. Kitaura et al. (2013) mention that apothecia of *L. fusisporum* “lack subhymenial paraplectechyma tissue, but have a paraplectenchymatous multilayered amphithecium instead”, but do not provide details of the source of this information. In any case, none of the combination of features described above are concurrent with those of *L. bellum* and *L. tiinae*.

**Taxonomy**

*Leptogium bellum* Kaasalainen sp. nov. (Fig. 1)

Mycobank # MB848314

Species of *Leptogium* with a plicate thallus, delicate nodules on thallus and apothecial thalline exciple, apothecia with a paraplectenchymatous proper exciple and one-celled cortex of the thalline exciple, and 40–60 µm long transversely septate ascospores.

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**TAXONOMY**

*Leptogium bellum* Kaasalainen sp. nov. (Fig. 1)

Mycobank # MB848314

Species of *Leptogium* with a plicate thallus, delicate nodules on thallus and apothecial thalline exciple, apothecia with a paraplectenchymatous proper exciple and one-celled cortex of the thalline exciple, and 40–60 µm long transversely septate ascospores.
Holotype: Tanzania, Mt. Kilimanjaro, upper montane secondary forest with *Erica excelsa* near the Maua Route, 3.1864° S 37.4403° E, 2820 m elevation, on a fallen branch, 11 March 2017, U. Kaasalainen UK170821a (H 9234932).

ITS barcoding marker accession (GenBank): MW340331 (holotype).

Description: Thallus up to 3 cm wide, opaque, glossy. Lobes laciniate, up to 5 mm wide, adnate or ascending. Upper surface longitudinally plicate, dirty white to gray; abundant, gray, delicate sack-like nodules clustered on lobe margins and apices, thallus ridges and the apothecial thalline exciple. Lower surface longitudinally plicate, dirty white or brownish grey. Isidia and lobules absent, but sack-like nodules on thallus and apothecia may resemble isidia. Rhizines and hairs absent, thallus attached to substrate by hapters. Apothecia common, up to 4.4 mm in diam., developing into the apices of stipe-like ascending lobes; discs of mature apothecia plane; thalline exciple gray, with abundant delicate nodules.

Anatomy: Thallus 200–450 μm thick, clearly thicker at ridges; quadratic cortical cells ca. 5 μm wide; medulla loose, columnar hyphae absent. Cyanobacteria (*Nostoc*) as long filaments, cells oval, 3–7 × 3–4 μm, heterocysts slightly larger. Apothecia with hymenium 110–140 μm thick; subhymenium 50–90 μm thick, orange-brownish; proper exciple paraplectenchymatous, 80–150 μm thick (12–15 cell layers) at the center, mainly pinkish grey, becoming more hyaline towards the (sub)hymenium; parahymenial tissue mainly consisting of the paraplectenchymatous proper exciple, 20–40(60) μm at the margin (on the level of hymenium surface); cortex of thalline exciple one cell layer thick. Ascospores acicular with 6–9 transverse septa, 40–60 × 6–12 μm (m = 52 × 9, SD = 6 × 1.7, N = 27). Pycnidia common and abundant, on and between the sack-like nodules, brown to dark brown; conidia rod-shaped, often slightly narrower at the middle, 3.4–4.0 × 1.5–1.8 μm in size.

Etymology: The specific epithet refers to the elegant appearance of the species.

Ecology and distribution: *Leptogium bellum* is so far only known from Mt. Kilimanjaro, Tanzania, where it is quite consistently present especially in middle and upper montane forests but also in subalpine shrublands (2260–3520 m). The species grows as an epiphyte on branches of montane trees and, in more open habitats in the subalpine zone and upper montane secondary *Erica excelsa* forest, also on tree trunks.

Notes: *Leptogium bellum* is easily distinguished from other *Leptogium* species with transversely septate ascospores due to the abundant, delicate, sack-like nodules on the thallus and thalline exciple. Additionally, the two species in East Africa, *L. bellum* and *L. tiinae*, are distinguished by ascospore size and habitat. *Leptogium bellum* occurs at high elevations (>2000 m) from middle montane forests to the subalpine zone and has shorter ascospores less than 60 μm long, while *L. tiinae* has ascospores over 60 μm long and is found from lower elevations (<1500 m) in relatively open habitats.

Additional specimens examined: Tanzania, Mt. Kilimanjaro, middle montane *Ocotea* forest near the Machame route, 3.1368° S 37.2455° E, 2260 m elevation, on a fallen branch, 9 March 2017, U. Kaasalainen UK170792g (H 9234931; ITS: MW340330, mtSSU: MW335316); near the Umbwe route, 3.1385° S 37.3052° E, 2650 m elevation, on a fallen branch, 20 June 2017, U. Kaasalainen UK171478e (H 9234938; ITS: MW340336, mtSSU: MW335322); 3.1400° S 37.3031° E, 2540 m elevation, on a fallen branch, 20 June 2017, U. Kaasalainen UK171516y (H 9234942; ITS: MW340339, mtSSU: MW335324); previously logged middle montane *Ocotea* forest near the Marangu route, 3.2018° S 37.5159° E, 2800 m elevation, on a fallen branch, 14 March 2017, U. Kaasalainen UK170880n (H 9234943; ITS: MW340332, mtSSU: MW335325); upper montane *Podocarpus* forest near the Mweka route, 3.1788° S 37.5126° E, 2940 m elevation, on a fallen branch, 24 June 2017, U. Kaasalainen UK171525k (H 9234940; ITS: MW340339, mtSSU: MW335325); upper montane secondary forest with *Erica excelsa* near the Maua route, 3.1864° S 37.4403° E,
2820 m elevation, on a fallen branch, 11 March 2017, U. Kaasalainen UK170822e (H 9234933); near the Mweka route, 3.1640° S 37.3675° E, 2990 m elevation, on a tree trunk, 25 June 2017, U. Kaasalainen UK171584c (H 9234941; ITS: MW340340, mtSSU: MW335326); subalpine Erica trimera forest near the Umbwe route, 3.1073° S 37.3184° E, 3510 m elevation, on an Erica trunk, 19 June 2017, U. Kaasalainen UK171439e (H 9234937; ITS: MW340335, mtSSU: MW335321); subalpine fire disturbed Erica shrubbery near the Machame route, 3.0854° S 37.2794° E, 3520 m elevation, on a branch in an Erica thicket, 15 June 2017, U. Kaasalainen UK171340j (H 9234936; ITS: MW340334, mtSSU: MW335320).

**Fig. 1. Leptogium bellum.** A–B. Type UK170821. A. General habit of moist thallus. B. Pycnidia on dry thallus. C. Habit when dry, showing the plicate thallus and delicate nodules on thallus and apothecia (UK171494b). D. Cross-section of an apothecium, showing the nodular thalline margin with one cell layer thick cortex and thick, paraplectenchymatous proper exciple (UK170911a). E. A transversely septate ascospore (UK170822e). Scales 2 mm in A and C, 200 µm in B and D, 10 µm in E.

**LEPTOGIUM TINAE** Kaasalainen & Rikkinen sp. nov. (Fig. 2)

Mycobank # MB848315

Species of *Leptogium* with a plicate thallus, robust nodules forming cerebroid structures on the apothecial thalline exciple and lobe apices, apothecia with a paraplectenchymatous proper exciple, one celled cortex of thalline exciple, and 60–90 µm long transversely septate ascospores.

Holotype: Kenya, Taita Taveta County, Wundanyi, alongside a dusty dirt road near the Taita Research Station of the University of Helsinki, 3.3981° S 38.3655° E, 1350 m elevation, on a basal trunk of large Ficus thonningii, 8 January 2010, J. Rikkinen JR10017 (EA; duplicate in H ─ H 9234943).
ITS barcoding marker accession (GenBank): JX503805 (holotype).

Description: Thallus up to 4 cm broad, opaque, usually glossy. Lobes laciniate, up to 3 mm wide, adnate, apices ascending, with robust and rounded cerebroid structures. Upper surface densely longitudinally plicate and wrinkled, dirty white to gray. Lower side longitudinally plicate, gray mottled with dirty white or yellow. Isidia and lobules absent. Cerebroid structures on thallus and apothecial margin formed by gray, wrinkled, sack-like nodules that become bloated when wet. Rhizines and hairs absent, thallus attached to substrate by hapters. Apothecia subpedicellate, up to 4.5 mm in diam., forming on the margins and apices of thallus lobes, often abundant and eventually covering a large portion of the thallus; disc in mature apothecia usually plane; thalline exciple gray, with abundant, robust and rounded cerebroid structures ca. 0.5−1.5 mm in diam.

Anatomy: Thallus 200−400 μm thick, several times thicker at the ridges; quadratic cortical cells 4−6 μm wide; medulla loose, columnar hyphae absent. Cyanobacteria (Nostoc) as long filaments, cells oval, 4−8 × 3−4 μm, heterocysts slightly larger. Apothecia with hymenium (95)130−170 μm thick; subhymenium (60)75−130 μm thick, orange-brownish; proper exciple paraplectenchymatous, (70)80−120 μm thick (8−10 cell layers) at the center, mainly pinkish grey, becoming more hyaline towards the (sub) hymenium; parahymenial tissue consisting mainly of the paraplectenchymatous proper exciple, 20−50 μm wide at the margin (on the level of hymenium surface); cortex of thalline exciple one cell layer thick. Ascospores acicular with 5−10(12) transverse septa, 60−90 × 5−9 μm (m = 70 × 7, SD = 9 × 0.9, N = 17). Pycnidia common and abundant, on and between the sack-like nodules, opening brown to black; conidia rod-shaped, often slightly

Fig. 2. Leptogium tiinae (JR10017 TYPE). A−B. Dry type material photographed in situ in the Taita Hills, showing the large and abundant apothecia with cerebroid structures on the thalline exciple. C. Pycnidia on cerebroid structures. D. Conidia from a pycnidium. E. Habit when moist. F. Apothecial section showing the paraplectenchymatous proper exciple. Scales 100 μm in C and F, 5 μm in D, 2 mm in E.
narrower in the middle, 3.6–4.1 × 1.3–1.7 µm in size.

Etymology: The specific epithet is dedicated to Estonian lichenologist Tiina Randlane, who visited the Taita Hills and the type locality of this species in October 2016.

Ecology and distribution: Fully developed specimens of *Leptogium tiinae* are known only from the type locality in the Taita Hills, Kenya. Additional specimens collected from a savanna habitat near Mt. Kilimanjaro, Tanzania, may represent *L. tiinae* (see Notes). All the specimens grew epiphytically on tree trunks in relatively open, sun exposed habitats at elevations below 1500 m.

Notes: The two specimens collected from Tanzania (UK160418b & d) are dark brown and notably smaller than the three well-developed thalli in the type collection of *L. tiinae*. Good quality ITS sequences were not obtained from these specimens, and their apothecia are not developed well enough to reveal all apothecial and ascospore characters. However, the mtSSU sequences confirm a close relationship between the two specimens and the type (Kaasalainen et al. 2021), and they are for now tentatively identified as *L. tiinae*.

*Leptogium tiinae* is easily distinguished from other *Leptogium* species with transversely septate ascospores based on the robust cerebroid structures that are especially abundant on the thalline exciple of the apothecia. For separation from *L. bellum*, see Notes under that species. Out of the previously described species with a similar apothecial anatomy, i.e., a paraplectenchymatous proper exciple and a one celled cortex of the thalline exciple, *L. tiinae* bears most resemblance to *L. longisporum*. However, the cerebroid structures on *L. tiinae* are more robust and rounded than those on *L. longisporum*. Additionally, the thallus of *L. tiinae* is thicker and always has abundant pycnidia, which have not been reported from *L. longisporum*.

Additional specimens examined: Tanzania, Kilimanjaro Region, Uchira, 3.3747° S 37.4549° E, 871 m elevation, on a trunk of a small tree on savanna, 10 December 2016, U. Kaasalainen UK16418b (H 9234944; mtSSU: MW335314), UK16418d (H 9234945; mtSSU: MW335315).

A key for *Leptogium* species with transversely septate ascospores

The key is based on inspected specimens (*L. bellum*, *L. chloromeloides*, *L. reticulatum*, *L. tiinae*), previously published descriptions (*L. ankolense*, *L. brebissonii*, *L. fusisporum*, *L. granadillae*, *L. longisporum*, *L. megapotamicum*, *L. pacificum*, *L. thoroldii*), or the combination of the two (*L. adpressum*). For *L. fusisporum*, both options for character combinations are included, following the descriptions given by Tuckerman (1882), Kitaura (2012), and Kitaura et al. (2013) and the description given by Sierk (1964). See Results and Discussion for more details and references.

1a. Thallus lobes large and robust, rounded, and clearly reticulately ridged – *Leptogium reticulatum*

1b. Lobes without reticulate ridges (but wrinkles and/or longitudinal ridges are often present) – 2.

2a. Lower cortex consisting of the basal cells of columnar hyphae connected by very thin longitudinal hyphae (pseudocortex) – *Leptogium pacificum*

2b. Lower cortex consisting of a single layer of ~isodiametric cells (eucortex) – 3.

3a. Apothecia with a thick paraplectenchymatous proper exciple (e.g. Figs 1D, 2F) – 4.

3b. Apothecia without a thick, paraplectenchymatous proper exciple – 10.

4a. Cortex of thalline exciple paraplectenchymatous, several cell layers thick – *Leptogium granadillae*

4b. Cortex of thalline exciple one cell layer thick (e.g. Fig. 1D) – 5.

5a. Thallus and apothecial margin with abundant delicate sack-like nodules (Fig. 1A, C) – *Leptogium bellum*
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REFERENCES


